

Effect of elevated CO₂ and temperature on thermal constants and lower threshold temperatures of maize aphid, *Rhopalosiphum maidis* (Fitch.) (Aphididae:Hemiptera) on maize, *Zea mays* (Linn.)

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ABSTRACT

The objective of this study was to examine the development of corn leaf aphid, *Rhopalosiphum maidis* Fitch (Aphididae: Hemiptera) on maize *Zea mays* Linnaeus at elevated and ambient concentrations of CO₂ (550 and 380ppm ± 25 ppm, respectively) at six temperatures (20, 25, 27, 30, 33 and 35°C) and to estimate thermal constants and lower temperature thresholds for the forecasting models based on heat accumulation units which could be developed for use in forecasting. The duration of different growth stages of *R. maidis* were reduced with an increase of temperature from 20°C to 35°C under both ambient and elevated CO₂ conditions. The lower development threshold for first nymphal instar, second nymphal instar, third nymphal instar, fourth nymphal instar, adult duration and total development period required 10.1, 5.04, 13.42, 26.96, 10.9, 23.22 and 20.20°C under eCO₂ whereas it was 13.32, 9.41, 19.13, 30.48, 16.38, 22.88 and 20.89°C under aCO₂ conditions, respectively. The mean lower temperature threshold for nymph was slightly higher (16.38°C) at aCO₂ compared to that of eCO₂ (10.90°C) whereas for adult the mean lower temperature threshold was slightly higher (23.22°C) at eCO₂ compared to that of aCO₂ (22.88°C). The thermal requirement of *R. maidis* from first nymphal instar to adult (total development period) was found to be 100.00 degree days (DD) under eCO₂ conditions as against 111.11 degree days under aCO₂ conditions. These estimated temperature thresholds and thermal constant can predict the pest scenarios and population dynamics of *R. maidis*.

Key words : *Rhopalosiphum maidis*, lower temperature threshold, thermal constant, degree days

Maize aphid *Rhopalosiphum maidis* (Fitch.) (Aphididae: Hemiptera), a sap-sucking homopteran insect is the largest group of phloem feeding insect. It is a polyphagous species occurring worldwide on sorghum, barley and wheat besides maize (Smith and Boyko, 2007). It is now distributed worldwide in the tropics and warmer temperate regions (Blackman and Eastop, 2000). Species of aphids are diverse and ubiquitous but their importance among crops differs (Srinivasa Rao *et al.*, 2016). Aphids cause mechanical harm and malnutrition to plants by the removal of phloem sap.

Agriculture is one of the most vulnerable sectors to the anticipated climate change with an adverse effect on crop yields. The increased levels of atmospheric CO₂ concentrations can have a direct effect on the growth rate of crop plants. Temperature has a direct influence on insect activity, rate of development and also plants. The predicted

changes in temperature and CO₂ concentration affect the population dynamics and the status of insect pests of various crops. Plants with C₄ photosynthesis will respond little to rising atmospheric CO₂ because a mechanism to increase the concentration of CO₂ in leaves causes CO₂ saturation of photosynthesis at ambient conditions. Numerous studies have shown that the annual average temperature of the earth will increase 1°C by 2025 and the probable rise in temperature by the end of the century is expected to reach 3°C (Solomon *et al.*, 2007). The parameters associated with life table are crucial for understanding population growth potential and for establishing effective management tactics to pest control in general because they provide information on development, reproduction and mortality of a pest population (Maia *et al.*, 2000).

So far, there is no published report from India on the effect of eCO₂ and variable temperatures on *R. maidis*.

Hence, in the present study, life table parameters of *R. maidis* were examined at two levels of CO₂ and six different temperatures to estimate the temperature thresholds and thermal constants which would be useful in status of the pest populations.

MATERIALS AND METHODS

Maintenance of Rhopalosiphum maidis culture

The corn leaf aphids, *R. maidis* were collected from the field and maintained at an optimum temperature of 27 ± 1°C and 75 ± 5% RH in the insectary of Entomology section, CRIDA, Hyderabad. The nymphs and adults were reared individually in petridishes (110 X 10 mm) to obtain bulk population for experiments. Light intensity of 30, 000 Lx was provided by 26 W florescent bulb inside the chambers during the 14 hours light period with a relative humidity of 60 % (day) and 70 % (night). Light illumination is provided through fluorescent lamps horizontally mounted in pairs above each shelf. Air circulation inside the chamber was maintained from a specifically designed air diffuser. The period of light, CO₂ concentrations and temperature levels were automatically monitored and controlled using Intellus Ultra Controller. The maize plants (DHM-117) and insects were maintained in open top chambers (OTC) and CO₂ growth chambers under respective set conditions at elevated and ambient concentrations of CO₂ (550 and 380ppm ± 25 ppm, respectively) at six temperatures of 20, 25, 27, 30, 33 and 35±1°C. Fully grown foliage (30 days after sowing) obtained from respective treatments was used for feeding trials and leaf quality analysis. The crop was maintained at insecticide free condition throughout the experiment to understand the impact of eCO₂ and temperature on insect pests. Experiments on life-tables of *R. maidis* were conducted by adopting cut leaf method at elevated and ambient concentrations of CO₂ (550 and 380ppm ±25 ppm, respectively) at six temperatures of 20, 25, 27, 30, 33 and 35±1°C and a photoperiod of 14L:10D. The cutcorn leaf (6x6 cm) from the top of corn seedlings of 1-2 months old was detached and placed in a petridish with a moist cotton swab on one side of the leaf margin. The cotton swabs were moistened daily to keep the leaf fresh and the leaves were changed on alternate days (Srinivasa Rao *et al.*, 2013). In order to construct life-tables, newly hatched 50 first instar nymphs were collected carefully from the stock culture with the help of wet camel hair brush and transferred individually into each petridish containing maize leaves obtained from respective set conditions with 50 replications per each

treatment. The data on durations of first instar, second instar, third instar, fourth instar, adult and total development period at each temperature under eCO₂ and aCO₂ conditions were recorded.

Calculation of lower temperature thresholds and thermal constants

Quantification of the relationship between insect development and temperature is useful to predict the seasonal occurrence and population dynamics of the insects. The degree day model (thermal summation model) was used to estimate the linear relationship between temperature and the rate of development of insect pests (Campbell *et al.*, 1974). The reciprocal of developmental period for each stage was calculated to obtain the development rate (1/day) at each temperature. Development threshold and thermal constant were determined by regressing development rate on temperature. Thermal constant is estimated by the formula Thermal constant (K) = (T – T₀) x D;

where, where, T - Temperature at which insect species is reared,

T₀ - Development threshold temperature,

D - Duration of development,

K was estimated as reciprocal of regression coefficient (b) between development rate and temperature. K = 1/ b.

To was determined as ratio of regression intercept 'a' and 'b' T₀ = -a/b. The life table parameters were estimated for eCO₂ and aCO₂ separately at abovementioned six temperatures and were plotted against the tested temperatures to compare the lower threshold temperatures and thermal constant under both eCO₂ and aCO₂ conditions.

RESULTS AND DISCUSSION

The durations of different growth stages *R. maidis* were significantly affected by eCO₂ and temperature conditions. The results pertaining to the variation in durations of first instar, second instar, third instar, fourth instar, adult and total development periods of *R. maidis* on maize at elevated and ambient concentrations of CO₂ (550 and 380ppm ± 25 ppm, respectively) at six different temperatures of 20, 25, 27, 30, 33 and 35±1°C are presented in Table 1.

The first nymphal instar duration (3.00 to 1.00 days), second nymphal instar (1.92 to 1.00 days), third nymphal instar (1.08 to 1.00 days), fourth nymphal instar (1.00 day), adult duration (23.1 to 1.20 days) and total development

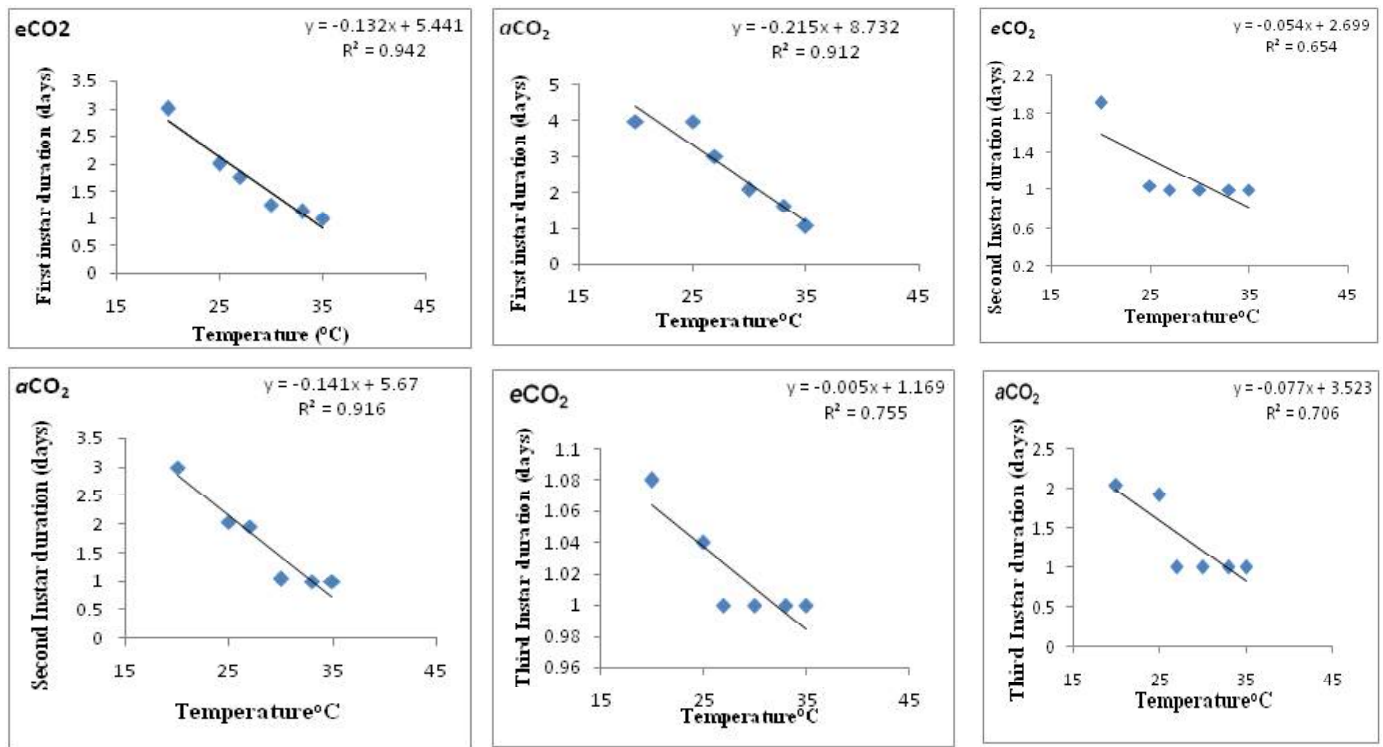


Fig. 1a: Effect of elevated CO₂ and temperatures on durations of first instar, second instar and third instar (days) of *R. maidis* on maize

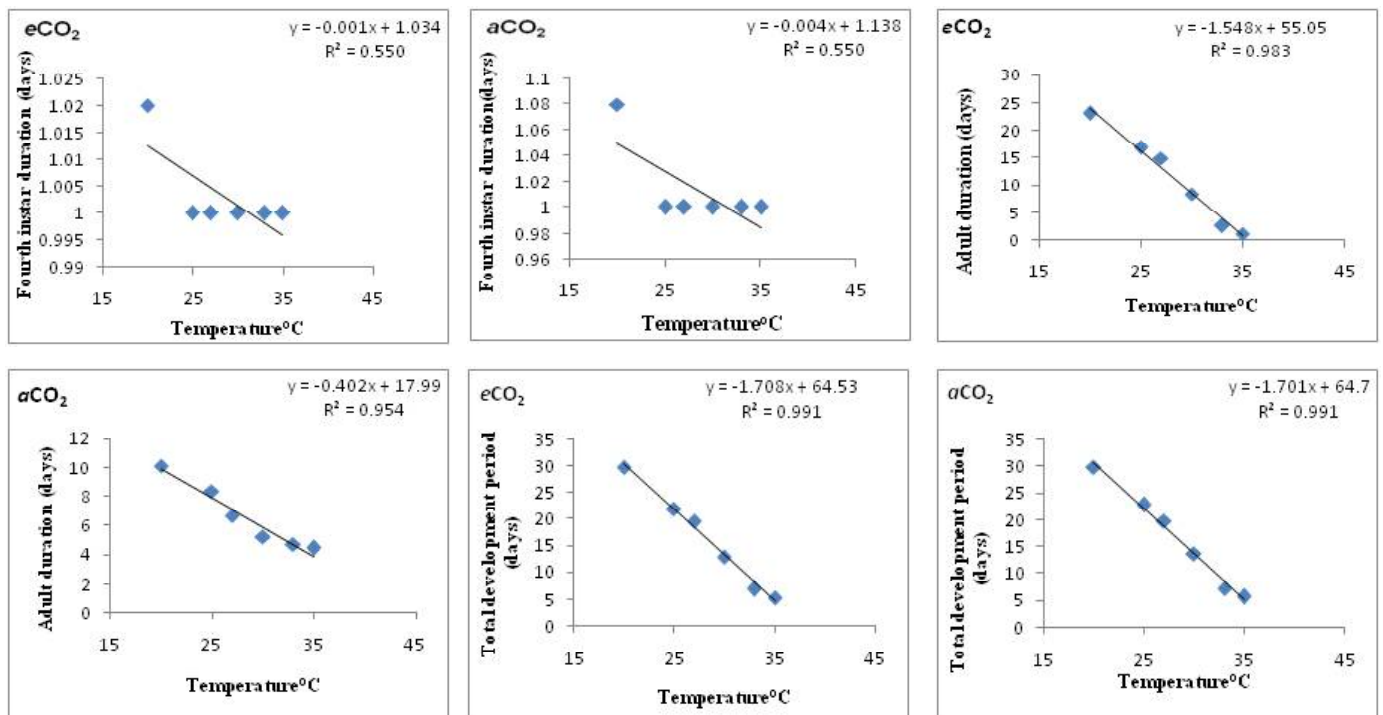


Fig 1b: Effect of elevated CO₂ and temperatures on fourth instar, adult duration and total development period (days) of *R. maidis* on maize

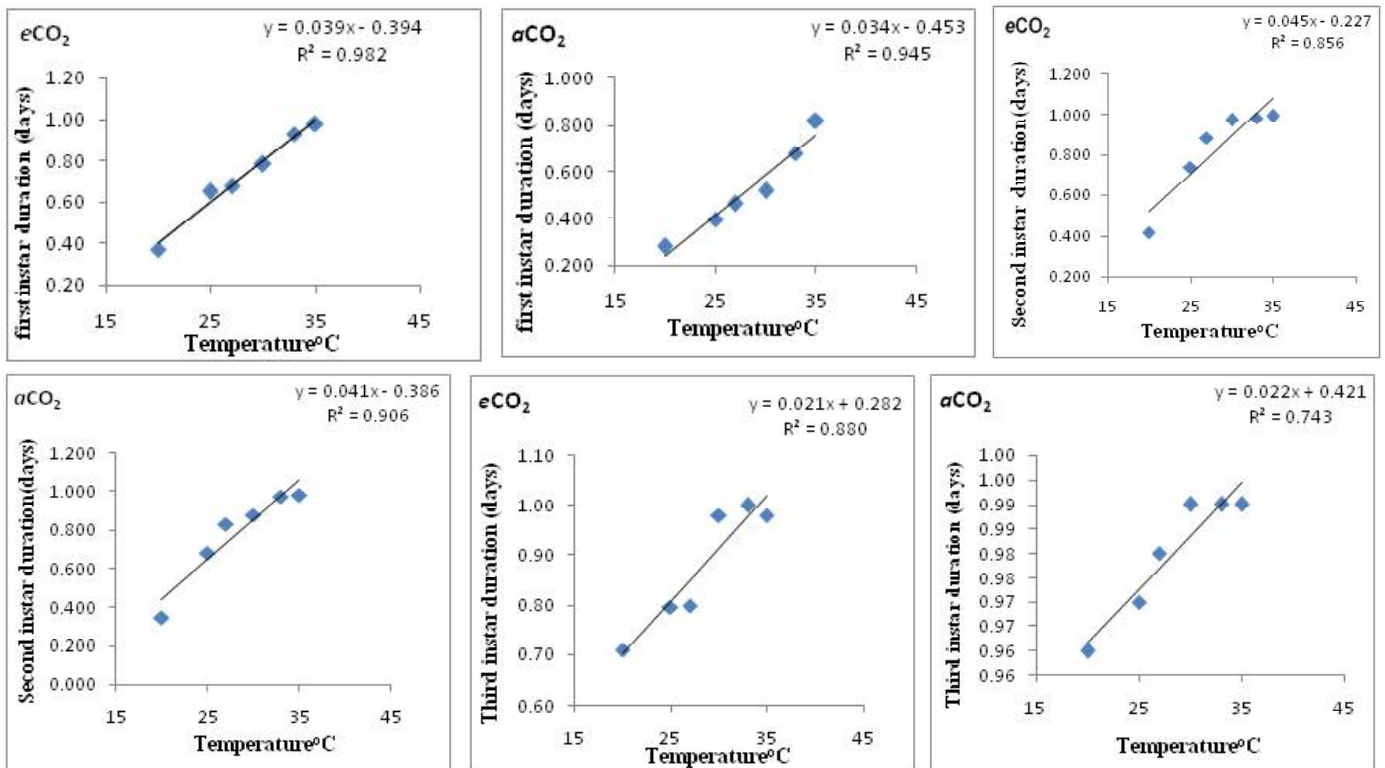


Fig.2a: Regression between temperatures and development rate to determine lower temperature threshold and thermal constant for durations of first instar, second instar and third instar duration (days) of *R. maidis* on maize under eCO_2 and aCO_2 conditions

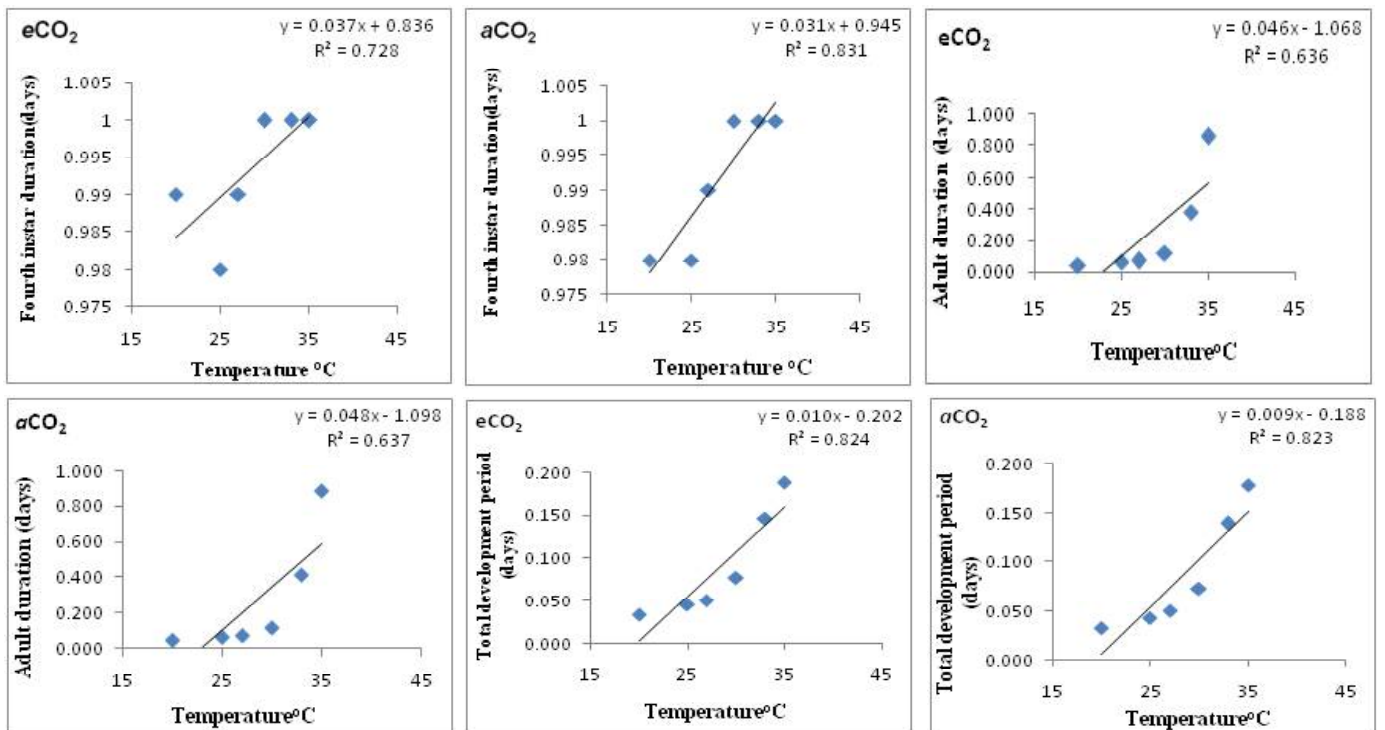


Fig. 2b: Regression between temperatures and development rate to determine lower temperature threshold and thermal constant for durations of fourth instar, adult duration and total development period (days) of *R. maidis* on maize

Table 1: Effect of elevated CO₂ and temperature on duration of different growth stages of *R. maidis* on maize

| Temperatures (°C) | First instar duration (days) | | Second instar duration (days) | | Third instar duration (days) | | Fourth instar duration (days) | | Adult duration (days) | | Total development period (days) | | |
|-----------------------------|------------------------------|---------------------------|-------------------------------|---------------------------|------------------------------|---------------------------|-------------------------------|---------------------------|---------------------------|---------------------------|---------------------------------|---------------------------|------------|
| | aCO ₂ (380ppm) | eCO ₂ (550ppm) | aCO ₂ (380ppm) | eCO ₂ (550ppm) | aCO ₂ (380ppm) | eCO ₂ (550ppm) | aCO ₂ (380ppm) | eCO ₂ (550ppm) | aCO ₂ (380ppm) | eCO ₂ (550ppm) | aCO ₂ (380ppm) | eCO ₂ (550ppm) | |
| 20 | 4.00±0.00 | 3.00±0.00 | 3.00±0.00 | 1.92±0.27 | 2.04±0.20 | 1.08±0.27 | 1.08±0.27 | 1.00±0.00 | 1.00±0.00 | 19.80±0.40 | 23.10±0.30 | 29.94±0.54 | 29.84±0.47 |
| 25 | 4.00±0.00 | 2.00±0.00 | 2.04±0.20 | 1.04±0.20 | 1.92±0.27 | 1.04±0.20 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 14.50±0.50 | 16.86±0.35 | 22.82±0.77 | 22.00±0.53 |
| 27 | 3.00±0.00 | 1.76±0.43 | 1.96±0.27 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 13.00±0.47 | 14.90±0.35 | 19.68±0.68 | 19.64±0.52 |
| 30 | 2.08±0.27 | 1.24±0.40 | 1.04±0.20 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 8.40±0.50 | 8.50±0.50 | 13.66±0.65 | 13.00±0.74 |
| 33 | 1.64±0.48 | 1.12±0.33 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 2.60±0.46 | 2.70±0.60 | 7.26±0.72 | 6.90±0.58 |
| 35 | 1.04±0.20 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.00±0.00 | 1.20±0.45 | 1.30±0.42 | 5.66±0.62 | 5.36±0.52 |
| F test | 80.44** | 121.24** | 145.51** | 2.09** | 242.73** | 5.98** | | | | | | | |
| S.Em± | 0.075 | 0.046 | 0.039 | 0.023 | 0.092 | 0.123 | | | | | | | |
| LSD(p=0.05) | 0.148 | 0.092 | 0.078 | 0.045 | 0.182 | 0.243 | | | | | | | |
| LSD(p=0.01) | 0.197 | 0.122 | 0.103 | 0.060 | 0.241 | 0.321 | | | | | | | |
| CV(%) | 12.30 | 11.9 | 11.88 | 7.94 | 4.27 | 3.81 | | | | | | | |
| Factor 1 (CO ₂) | | | | | | | | | | | | | |
| aCO ₂ (380 ppm) | 2.620 | 1.67 | 1.32 | 1.013 | 9.91 | 16.50 | | | | | | | |
| eCO ₂ (550 ppm) | 1.680 | 1.18 | 1.02 | 1.000 | 11.21 | 16.12 | | | | | | | |
| F test | 965.88** | 1059.87** | 378.99** | 2.09NS | 959.34** | 60.86** | | | | | | | |
| S.Em± | 0.030 | 0.015 | 0.016 | 0.009 | 0.042 | 0.048 | | | | | | | |
| LSD(p=0.05) | 0.062 | 0.031 | 0.033 | NS | 0.084 | 0.097 | | | | | | | |
| LSD(p=0.01) | 0.085 | 0.042 | 0.044 | NS | 0.112 | 0.129 | | | | | | | |
| CV(%) | 12.18 | 9.20 | 11.63 | 7.94 | 4.85 | 3.62 | | | | | | | |
| Factor 2 (Temperature(°C)) | | | | | | | | | | | | | |
| 20 | 3.50 | 2.46 | 1.56 | 1.04 | 21.45 | 29.89 | | | | | | | |
| 25 | 3.00 | 1.60 | 1.48 | 1.00 | 15.67 | 22.41 | | | | | | | |
| 27 | 2.36 | 1.48 | 1.00 | 1.00 | 13.94 | 19.66 | | | | | | | |
| 30 | 1.64 | 1.02 | 1.00 | 1.00 | 8.45 | 13.34 | | | | | | | |
| 33 | 1.38 | 1.00 | 1.00 | 1.00 | 2.63 | 7.08 | | | | | | | |
| 35 | 1.02 | 1.00 | 1.00 | 1.00 | 1.25 | 5.51 | | | | | | | |
| F. test | 675.21** | 563.19** | 187.06** | 2.09** | 30482.27** | 22952.31** | | | | | | | |
| S.Em± | 0.053 | 0.034 | 0.028 | 0.016 | 0.064 | 0.088 | | | | | | | |
| LSD(p=0.05) | 0.104 | 0.067 | 0.055 | 0.032 | 0.125 | 0.173 | | | | | | | |
| LSD(p=0.01) | 0.137 | 0.088 | 0.073 | 0.042 ^c | 0.165 | 0.228 | | | | | | | |

All values are mean ± standard deviation;

** Significant @ 1% level of significance NS = Not-significant

Table 2: Effect of elevated CO₂ and temperature on lower temperature thresholds and thermal constants of each developmental stage of *R. maidis* on maize

| Development stage | Regression equation | | R ² value | | Lower temperature Threshold (T ₀ =a/b) (°C) | | Thermal constant (K=l/b) (Degree Days-DD) | |
|--------------------------|-----------------------------------|------------------------------------|-----------------------------------|------------------------------------|--|------------------------------------|---|------------------------------------|
| | Ambient CO ₂ (380 ppm) | Elevated CO ₂ (550 ppm) | Ambient CO ₂ (380 ppm) | Elevated CO ₂ (550 ppm) | Ambient CO ₂ (380 ppm) | Elevated CO ₂ (550 ppm) | Ambient CO ₂ (380 ppm) | Elevated CO ₂ (550 ppm) |
| | First instar | y = 0.034x - 0.453 | y = 0.039x - 0.394 | 0.945 | 0.982 | 13.32 | 10.10 | 29.41 |
| Second instar | y = 0.041x - 0.386 | y = 0.045x - 0.227 | 0.906 | 0.856 | 9.41 | 5.04 | 24.39 | 22.22 |
| Third instar | y = 0.022x + 0.421 | y = 0.021x + 0.282 | 0.743 | 0.880 | 19.13 | 13.42 | 45.45 | 41.66 |
| Fourth instar | y = 0.031x + 0.945 | y = 0.037x + 0.836 | 0.831 | 0.728 | 30.48 | 26.96 | 32.25 | 27.02 |
| Nymphal duration | y = 0.008x + 0.131 | y = 0.009x - 0.098 | 0.962 | 0.966 | 16.38 | 10.9 | 125.00 | 111.11 |
| Adult | y = 0.046x - 1.068 | y = 0.048x - 1.098 | 0.637 | 0.636 | 22.88 | 23.22 | 20.83 | 21.74 |
| Total development period | y = 0.009x - 0.188 | y = 0.010x - 0.202 | 0.823 | 0.824 | 20.89 | 20.20 | 111.11 | 100.00 |

period (29.84 to 5.36 days) were recorded under eCO₂ conditions. Similar trend of reduction of the first nymphal instar duration (4.00 to 1.04 days), second nymphal instar (3.00 to 1.00 days), third nymphal instar (2.04 to 1.00 days), fourth nymphal instar (1.08 to 1.00 days), adult duration (19.8 to 1.30 days) and total development period (29.94 to 5.66 days) were recorded under aCO₂ conditions (Fig 1a & 1b). The total development period was decreased with increasing temperature from 20 to 35°C under both eCO₂ and aCO₂ conditions. The present results are in conformity with the findings of Kuo *et al.* (2006), who reported that the duration of total immature stages of *R. maidis* from birth to adult decreased as temperature increased from 6°C (51.7 days) to 30°C (5.2 days). The total immature stages of *R. maidis* from birth to adult fed on barley and maize foliage, the estimated low developmental threshold (4.44°C and 6.1°C) and the shortest developmental time at 26°C and 30°C (Elliott *et al.*, 1988), respectively. Total immature developmental duration of *Nilaparvata lugens* decreased from 74.4 days at 15°C to 22.4 days at 30°C (Vattikuti *et al.*, 2019). This means that *R. maidis* have a wider temperature range for immature development on corn leaves than on barley leaves. The present results indicated that the lower degree days requirement for completion of nymphal stage at eCO₂ and might be due to shortening of development time.

Lower threshold temperatures and thermal constants of *R. maidis*

Linear regression model was used to calculate the lower temperature threshold and thermal constant. The development rate of each stage of insect was regressed on temperature and linear regression equations are depicted in the Table 2. The linear regression equations of development rate with temperature under eCO₂ conditions indicated significant multiple coefficient of determination for first nymphal instar (y = 0.039x - 0.394, R²= 0.982), second nymphal instar (y= 0.045x - 0.227, R²=0.856), third nymphal instar (y= 0.021x + 0.282, R²= 0.880), fourth nymphal instar (y = 0.037x + 0.836, R²= 0.728), total nymphal duration (y = 0.009x - 0.098, R²= 0.966), adult duration (y = 0.048x - 1.098, R²= 0.636) and total development period (y = 0.010x - 0.202, R²= 0.824). Similarly the linear regression equations for different stages of *R. maidis* under aCO₂ indicated significant multiple coefficient of determination for first nymphal instar (y = 0.034x - 0.453, R²= 0.945), second nymphal instar (y= 0.041x - 0.386, R²= 0.906), third nymphal instar (y = 0.022x + 0.421, R²= 0.743), fourth nymphal instar (y = 0.031x + 0.945, R²= 0.831), total nymphal duration (y

= 0.008x + 0.131, R²= 0.962), adult duration ($y = 0.046x - 1.068$, R²= 0.637) and total development period ($y = 0.009x - 0.188$, R²= 0.823) (Fig 2a & 2b). Development rate and temperature were separately plotted to determine the lower development threshold of *R. maidis* at two CO₂ conditions. The lower development threshold for durations of first, second, third, fourth nymphal instar, total nymphal duration, adult duration and total development period required 10.1, 5.04, 13.42, 26.96, 10.9, 23.22 and 20.20°C under eCO₂ whereas it was 13.32, 9.41, 19.13, 30.48, 16.38, 22.88 and 20.89°C under aCO₂ conditions, respectively. The mean lower development threshold for nymph was slightly higher (16.38°C) at aCO₂ compared to that of eCO₂ (10.90°C) whereas for adult the mean lower development threshold was slightly higher (23.22°C) at eCO₂ compared to that of aCO₂ (22.88°C). The rate of insect development is affected by the temperature to which insects are exposed. The thermal requirement of *R. maidis* from first nymphal instar to adult (total development period) was found to be 100.00 degree days (DD) under eCO₂ conditions as against 111.11 degree days under aCO₂ conditions. Durations of first, second, third and fourth nymphal instar, total nymphal duration, adult duration required 25.64, 22.22, 41.66, 27.02, 111.11 and 21.74 DD under eCO₂ conditions as against 29.41, 24.39, 45.45, 32.25, 125.00 and 20.83 DD under aCO₂ conditions, respectively. Thus, the thermal requirements indicated that the lower DD requirement for completion of nymphal stage at eCO₂ compared to that of aCO₂. In contrast, adult required higher DD at eCO₂ with the corresponding temperatures of 20-35°C (Table 2).

According to lower temperature thresholds and thermal constants, the results were in conformity with the findings of Kuo *et al.* (2006) who reported that the lower thermal constants for *R. maidis* from 33.15 DD for first instars to 25.96 DD for fourth instars. Similar results were observed by Srinivasa Rao *et al.* (2017) who reported that the thermal requirement of nymph varied from 74-102 DD on eCO₂ with temperature in the range of 20-30°C as against 90-130 DD on aCO₂ and the results indicated that the lower degree days requirement for completion of nymphal stage at eCO₂. The lower thermal constants of *A. craccivora* at eCO₂ conditions might be due to reduction of DT as reported earlier (Berberet *et al.*, 2009). Similar kind of variation of 513-290 DD for soybean aphid was reported by McCornack *et al.* (2004) in the 20-35°C temperature range. These lower thermal requirements might be due to feeding on low nutritious maize foliage obtained from eCO₂ conditions. It was suggested that the thermal constant is influenced not

only by the temperature but also by the biochemical constituents present in the host plant viz., nitrogen, proteins, aminoacids and tannins (Marchioro and Foerster, 2011). The estimated temperature thresholds and thermal constants are useful in the prediction of population peaks (Taveras *et al.*, 2004) and the present data will be relevant in future climate change scenarios to understand the distribution and abundance of insect pest.

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